

Evidence for the bone structure change and osteocytes' biorhythm during orthodontic tooth movement



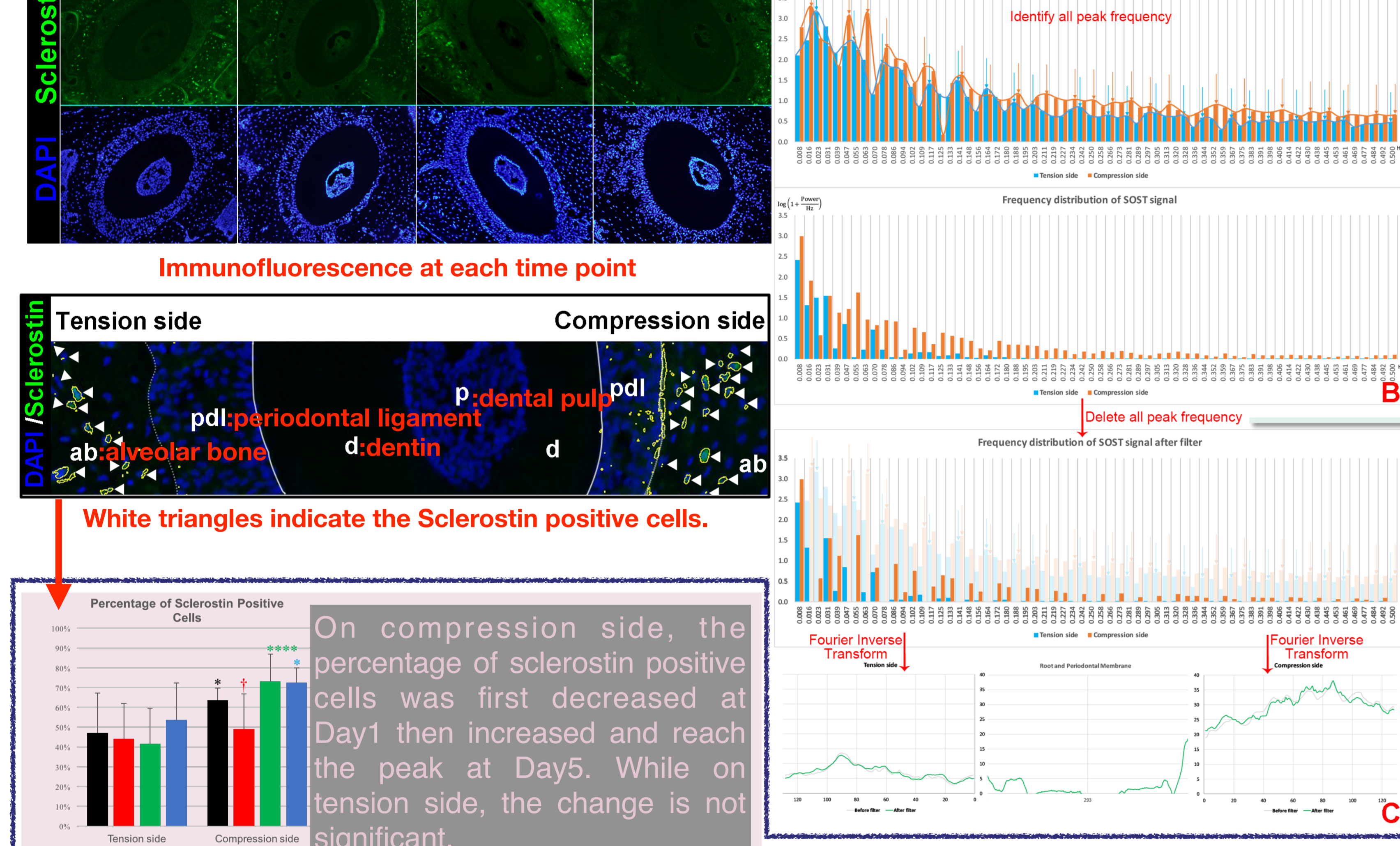
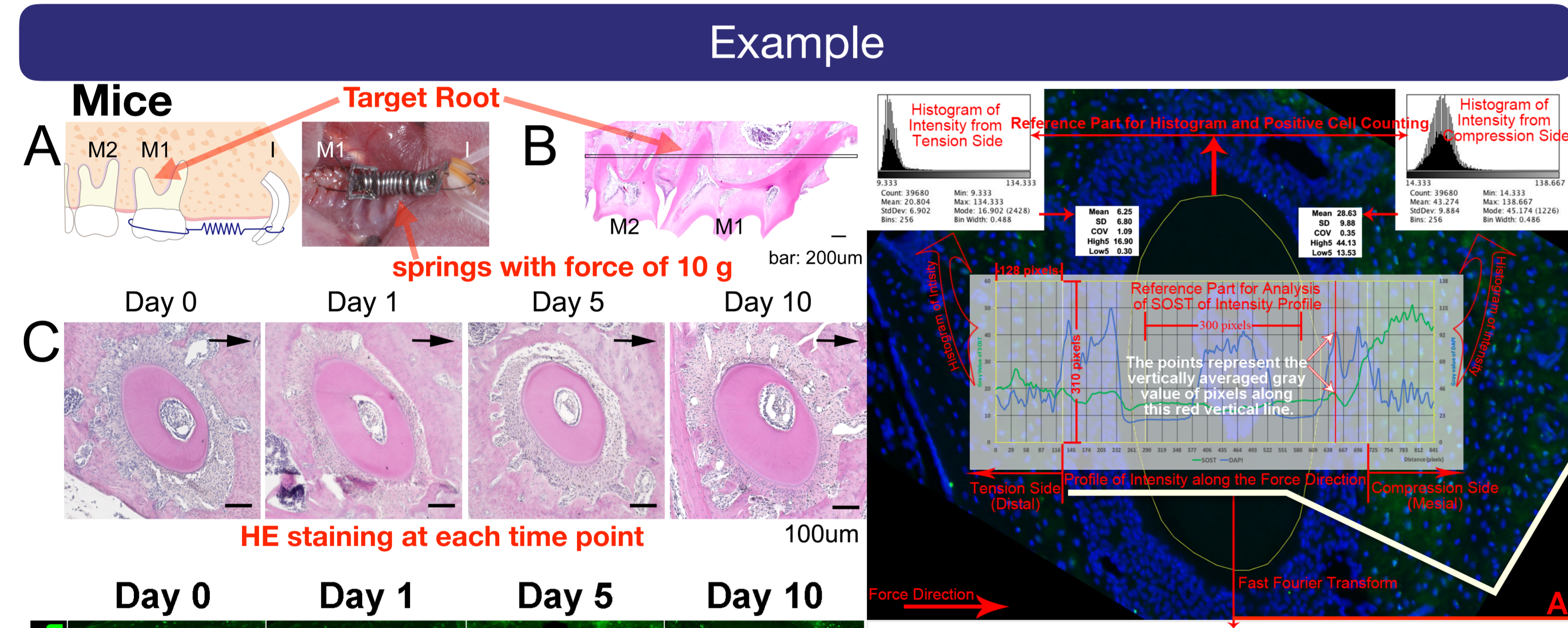
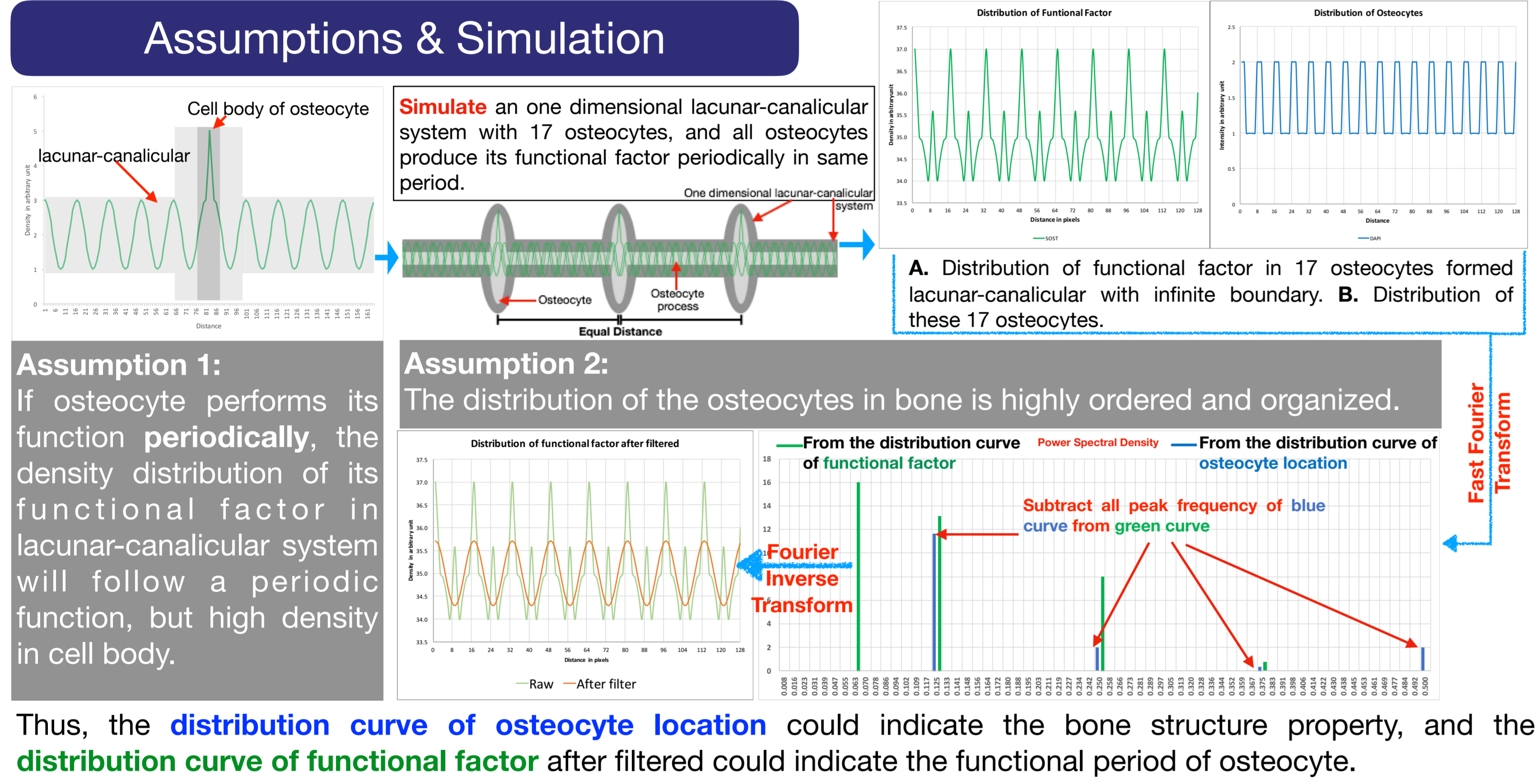
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Introduction

The **objective** of this study is to investigate the relationship between the change of sclerostin spatial distribution and the morphological change of bone structure during orthodontic tooth movement. **Sclerostin** is the key regulator of bone modeling and remodeling, which is almost exclusively formed by osteocytes and has anti-anabolic effects on bone formation. Meanwhile, as the most plenty cell emerged in bone matrix, **osteocytes** is an appropriate indicator for the property of bone structure. In this study, we used **fast Fourier transform (FFT)** and **wavelet transform** to reveal the tiny changes in bone structure and to detect the changes of sclerostin spatial distribution.

Materials and Methods



Spearman's rho

Table 1. Significant relative factors^a (34 sections from 18 mice)

Factors	Spearman's correlation coefficient	P-Value
Factors relative to the percentage of sclerostin positive cells		
MPSDF of sclerostin signal	0.35	<0.00**
Mean	0.43	<0.00***
SD	0.36	<0.00**
COV	-0.24	<0.05*
High5	0.48	<0.00***
Factors relative to the MPSDF of DAPI signal		
High5	0.25	<0.05*
Factors relative to the MPSDF of sclerostin signal after filter		
Low5	-0.28	<0.03*

^a Mixed compression, tension side and every time point. MPF: mean power frequency. SOST: sclerostin.
 *P<0.05, **P<0.01, ***P<0.001, ****P<0.0001

Multiple linear regression

Table 2. The results of multiple linear regression^a (34 sections from 18 mice)

Factors	R ²	F statistic	P-Value	Coefficient estimate±SE	t value	P-Value	Square of semipartial correlations	VIF ^b
The percentage of sclerostin positive cells	0.38	13.067	<0.000***					
Intercept				36.12 ± 5.62	6.43	<0.000***		
The MPSDF of sclerostin signal				373.25 ± 153.92	2.43	<0.02*	0.06	1.06
Mean				1.61 ± 0.31	5.27	<0.000***	0.27	1.22
Low5				-1.66 ± 0.34	-3.43	<0.000***	0.11	1.29

^a VIF: variance inflation factor; ^b VIF higher than 10 suggests a linear relationship between the predictors; SE: standard error.
 *P<0.05, **P<0.01, ***P<0.001, ****P<0.0001

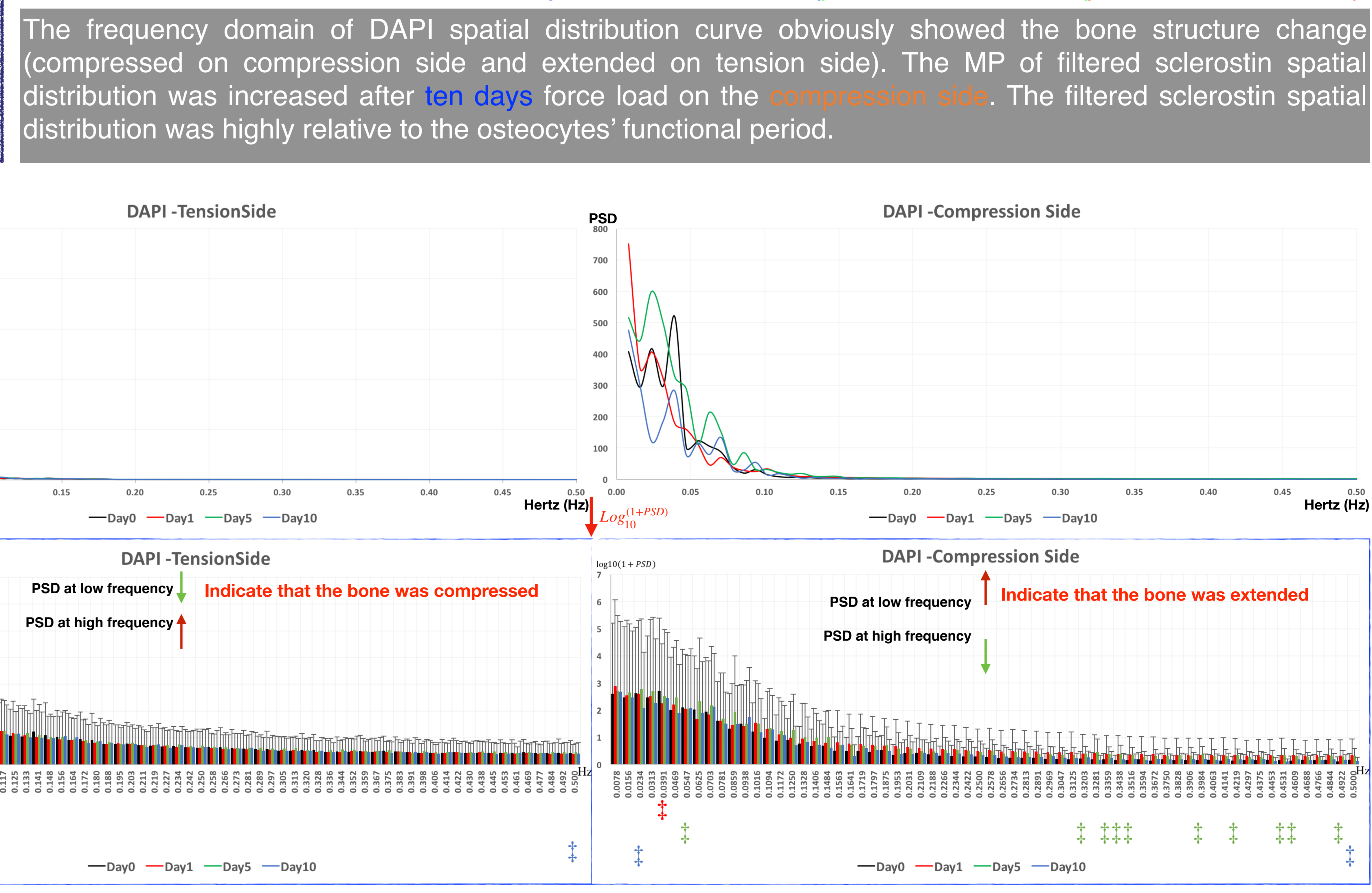
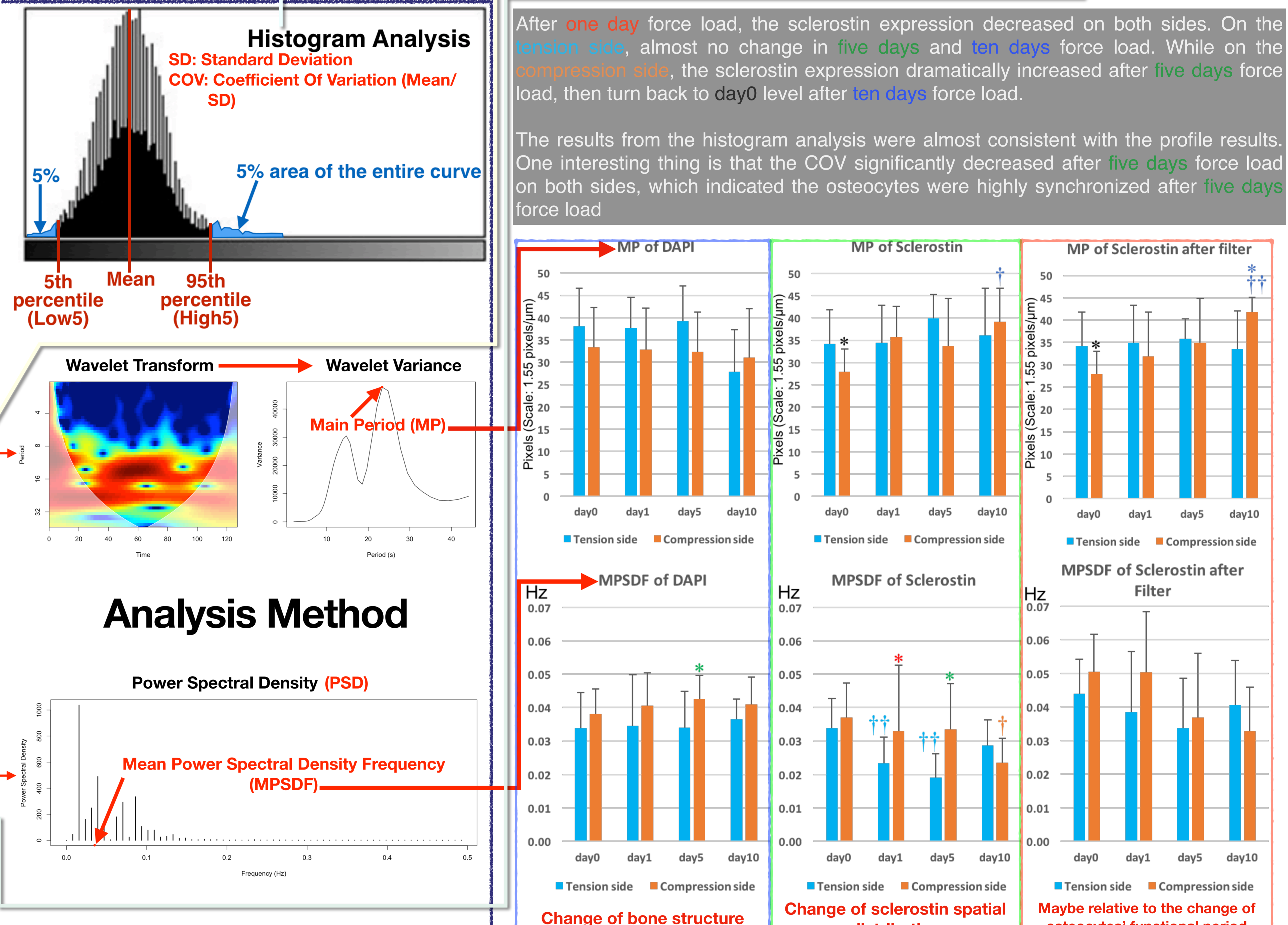
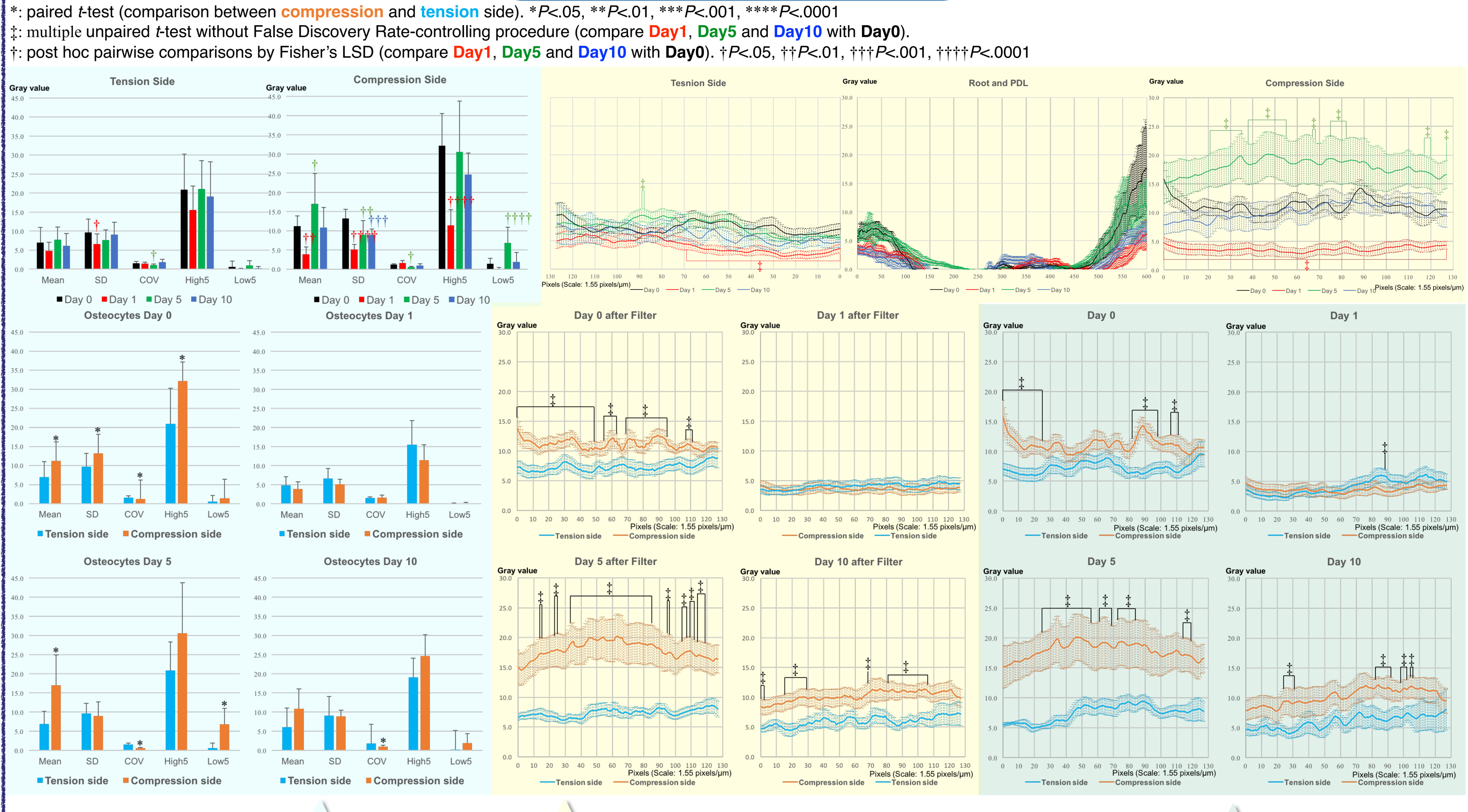
The High5 have the highest correlation coefficient to the percentage of sclerostin positive cells and have significant correlation to the MPSDF of DAPI signal (represent the bone structure property). This means the High5 cells can response to the bone structure change and impact the sclerostin expression. The MPSDF of sclerostin signal (indicate the period of sclerostin expression) also have correlation with the sclerostin positive cells.

Conclusions

- In this study:**
- We revealed the spatial distribution change of sclerostin expression during the orthodontic tooth movement.
 - The spatial distribution change of sclerostin expression have association with bone structure change.
 - The spatial distribution change of sclerostin expression maybe mainly induced by the osteocytes' functional period.

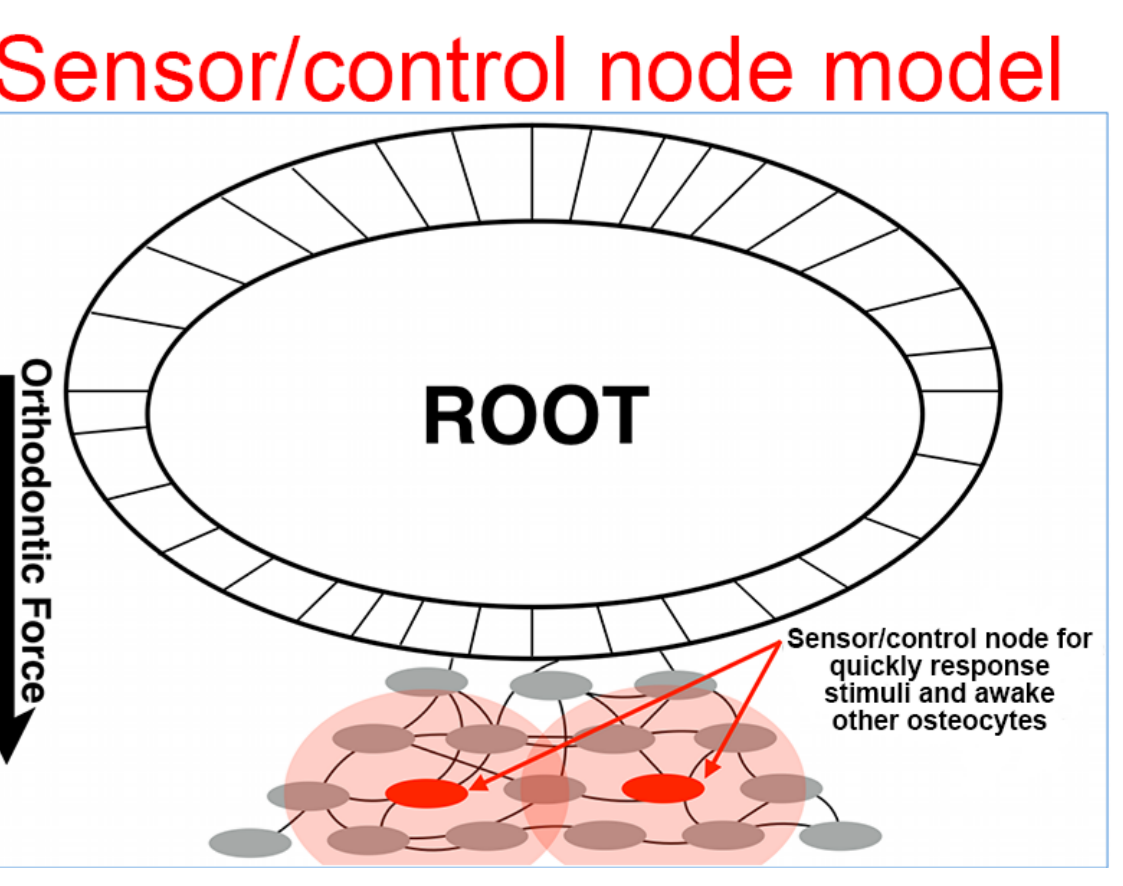
The Japanese Orthodontic Society COI Disclosure. Ziyi WANG, Yoshihito ISHIHARA, Naoya ODAGAKI, Masahiro NAKAMURA, Ei EI HSU HLAING, Hiroshi KAMIOKA. The authors have no financial conflicts of interest to disclose concerning the presentation.

Results



Hypothesis

Our previous study [1] showed that not all osteocytes have observable gap junctional intercellular communication (GJIC) in chick calvariae and demonstrated two types of GJICs in the development of mature osteocytes in chicks: passive transduction (low GJIC osteocyte) and active transduction (high GJIC osteocyte). Combination of the findings in this time, we hypothesize a pattern in which high5 osteocytes function as a sensor/control node that maintains a high GJIC to allow a quick response to stimuli, to activate the surrounding osteocytes, and to maintain the coordination of the period of the sclerostin expression.



Reference

1. Wang, Ziyi, et al. "Alteration in the gap-junctional intercellular communication capacity during the maturation of osteocytes in the embryonic chick calvaria." *Bone* 91 (2016): 20-29.